

Package ‘GENEAcore’

November 21, 2025

Title Pre-Processing of 'GENEActiv' Data
Version 1.1.2
Date 2025-11-21
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Description
Analytics to read in and segment raw 'GENEActiv' accelerometer data into epochs and events.
For more details on the 'GENEActiv' device, see <<https://activinsights.com/resources/geneactiv-support-1-2/>>.
License GPL (>= 2)
Encoding UTF-8
RoxygenNote 7.3.2
Imports changepoint, signal, methods
Suggests knitr, rmarkdown, testthat (>= 3.0.0), GENEAread (>= 2.0.10),
GENEAclassify
Config/testthat/edition 3
VignetteBuilder knitr
NeedsCompilation no
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Repository CRAN
Date/Publication 2025-11-21 17:20:02 UTC

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aggregate_epochs	<i>Aggregate Epochs</i>
------------------	-------------------------

Description

Aggregate Epochs

Usage

```
aggregate_epochs(  
  time_series,  
  measure = "AGSA",  
  time = "timestamp",  
  sample_frequency,  
  duration = NA,  
  first_epoch_timestamp = NA,  
  fun = mean  
)  
  
aggregateEpochs(...)
```

Arguments

time_series	Data frame to be aggregated.
measure	Name of the measure columns to be included.
time	Name of the time column.
sample_frequency	Measurement frequency of data.
duration	Time duration to aggregate in each epoch.
first_epoch_timestamp	Time to start the first epoch, defaults to first record.
fun	Function to apply on aggregation, defaults to mean.
...	Additional arguments passed to internal aggregation functions.

Details

Wrapper function that calls `aggregate_periods` for epochs (duration of fixed length).

Value

Data frame of aggregated epochs.

Examples

```
timestamp <- c(
  1619424004, 1619424005, 1619424006, 1619424007,
  1619424008, 1619424009, 1619424010, 1619424011,
  1619424012, 1619424013, 1619424014, 1619424015
)
value <- c(
  0.729614366, 1.729115871, 0.804973546, 2.510181118,
  2.23764038, 0.613203747, 0.681953275, 0.089566943,
  0.021042388, 2.4780338, 2.437488989, 2.632635727
)
data <- data.frame(timestamp, value)
aggregated <- aggregate_epochs(data,
  duration = 5,
  measure = "value",
  sample_frequency = 1,
  first_epoch_timestamp = 1619424005,
  time = "timestamp"
)
```

aggregate_events	<i>Aggregate Events</i>
------------------	-------------------------

Description

Aggregate Events

Usage

```
aggregate_events(
  time_series,
  measure = "AGSA",
  time = "timestamp",
  sample_frequency,
  events = NA,
  start_time = "start",
  end_time = "end",
  fun = mean
)

aggregateEvents(...)

aggregatePeriods(...)

createEventMapping(...)
```

Arguments

time_series	Data frame to be aggregated.
measure	Name of the measure columns to be included.
time	Name of the time column.
sample_frequency	Measurement frequency of data.
events	Data frame containing the start and end index of each event.
start_time	Name of the column in events containing the start index of the events.
end_time	Name of the column in events containing the end index of the events.
fun	Function to apply on aggregation, defaults to mean.
...	Additional arguments passed to internal aggregation functions.

Details

Wrapper function that calls aggregate_periods for events (duration of variable length).

Value

Data frame of aggregated events.

Examples

```

timestamp <- c(
  1619424004, 1619424005, 1619424006, 1619424007,
  1619424008, 1619424009, 1619424010, 1619424011,
  1619424012, 1619424013, 1619424014, 1619424015
)
value <- c(
  0.729614366, 1.729115871, 0.804973546, 2.510181118,
  2.23764038, 0.613203747, 0.681953275, 0.089566943,
  0.021042388, 2.4780338, 2.437488989, 2.632635727
)
data <- data.frame(timestamp, value)
event_start <- c(1, 5, 10)
event_end <- c(4, 9, 12)
aggregated_events <- aggregate_events(data,
  events = data.frame(start = event_start, end = event_end),
  measure = "value",
  time = "timestamp",
  start_time = "start",
  end_time = "end",
  sample_frequency = 1,
  fun = sum
)

```

aggregate_periods	<i>Aggregate Periods</i>
-------------------	--------------------------

Description

Generalised aggregation function generates distinct epochs or events outputs based on the initial parameters provided.

Usage

```

aggregate_periods(
  time_series,
  measure = "AGSA",
  time = "timestamp",
  sample_frequency,
  duration = NA,
  first_epoch_timestamp = NA,
  events = NA,
  start_time = "start",
  end_time = "end",
  fun = mean
)

```

Arguments

time_series	Data frame to be aggregated.
measure	Name of the measure columns to be included.
time	Name of the time column.
sample_frequency	Frequency of data.
duration	Time duration to aggregate in each epoch.
first_epoch_timestamp	Time to start the first epoch, defaults to first record.
events	Data frame containing the start and end index of each event.
start_time	Name of the column in events containing the start index of the events.
end_time	Name of the column in events containing the end index of the events.
fun	Function to apply on aggregation, defaults to mean.

Value

Data frame of aggregated epochs or events.

apply_AGSA	<i>Apply Absolute Gravity-Subtracted Acceleration (AGSA)</i>
------------	--

Description

Apply Absolute Gravity-Subtracted Acceleration (AGSA)

Usage

```
apply_AGSA(x)
```

Arguments

x	Calibrated acceleration data frame.
---	-------------------------------------

Value

Measure column appended to end of calibrated data frame.

Examples

```
x <- c(0.14268, 0.21757, -0.529, -0.36383)
y <- c(0.26385, 0.27295, 0.29220, 0.79510)
z <- c(0.27722, 0.20296, 0.35092, 0.27459)
calibrated <- data.frame(x, y, z)
calibrated <- apply_AGSA(calibrated)
```

apply_all	<i>Apply AGSA, ENMO, UpDown and Degrees</i>
-----------	---

Description

Single function that combines calculations for acceleration, elevation and rotation measures.

Usage

```
apply_all(x)
```

Arguments

x Calibrated acceleration data frame.

Value

Measures columns appended to end of calibrated data frame.

Examples

```
x <- c(0.14268, 0.21757, -0.529, -0.36383)
y <- c(0.26385, 0.27295, 0.29220, 0.79510)
z <- c(0.27722, 0.20296, 0.35092, 0.27459)
calibrated <- data.frame(x, y, z)
calibrated <- apply_all(calibrated)
```

apply_calibration	<i>Apply Calibration</i>
-------------------	--------------------------

Description

Apply Calibration

Usage

```
apply_calibration(sensor_data, cal_params, measurement_device, use_temp = TRUE)
```

Arguments

sensor_data	Raw sensor-level data from a bin file in the form (x, y, z, Light, Button, Temp).
cal_params	Calibration parameters for acceleration and light from MPI.
measurement_device	Name of the measurement device used "GENEActiv 1.1" or "GENEActiv 1.2".
use_temp	Allows auto-calibration to be run with and without temperature compensation.

Details

Function to apply calibration to sensor-level data from a bin file.

Value

Data frame of calibrated sensor data.

Examples

```
cal_params <- list(
  scale = c(1.015, 1.017, 1.027),
  offset = c(0.00128, 0.0383, 0.0138),
  temperature_offset = c(0, 0, 0),
  error = NA,
  light_denominator = 48,
  light_numerator = 911
)

rawdata <- data.frame(
  time = c(rep(1726650857, 5)),
  x = c(0.2421875, 0.24609375, 0.25390625, 0.24609375, 0.23828125),
  y = c(-0.04296875, -0.04687500, -0.03515625, -0.03125000, -0.04296875),
  z = c(-0.9453125, -0.9453125, -0.9531250, -0.9531250, -0.9609375),
  light = c(rep(22, 5)),
  button = c(rep(0, 5)),
  temp = c(rep(21.3, 5)),
  volts = c(rep(4.0896, 5))
)

calibrated <- apply_calibration(rawdata, cal_params, "GENEActiv 1.1")
```

apply_degrees	<i>Apply Rotation (Degrees)</i>
---------------	---------------------------------

Description

Apply Rotation (Degrees)

Usage

```
apply_degrees(x)
```

Arguments

x Calibrated acceleration data frame.

Value

Measure column appended to end of calibrated data frame.

Examples

```
x <- c(0.14268, 0.21757, -0.529, -0.36383)
y <- c(0.26385, 0.27295, 0.29220, 0.79510)
z <- c(0.27722, 0.20296, 0.35092, 0.27459)
calibrated <- data.frame(x, y, z)
calibrated <- apply_degrees(calibrated)
```

apply_ENMO	<i>Apply Euclidean Norm Minus One (ENMO)</i>
------------	--

Description

Apply Euclidean Norm Minus One (ENMO)

Usage

```
apply_ENMO(x)
```

Arguments

x Calibrated acceleration data frame.

Value

Measure column appended to end of calibrated data frame.

Examples

```
x <- c(0.14268, 0.21757, -0.529, -0.36383)
y <- c(0.26385, 0.27295, 0.29220, 0.79510)
z <- c(0.27722, 0.20296, 0.35092, 0.27459)
calibrated <- data.frame(x, y, z)
calibrated <- apply_ENMO(calibrated)
```

apply_radians	<i>Apply Rotation (radians)</i>
---------------	---------------------------------

Description

Apply Rotation (radians)

Usage

```
apply_radians(x)
```

Arguments

x Calibrated acceleration data frame.

Value

Measure column appended to end of calibrated data frame.

Examples

```
x <- c(0.14268, 0.21757, -0.529, -0.36383)
y <- c(0.26385, 0.27295, 0.29220, 0.79510)
z <- c(0.27722, 0.20296, 0.35092, 0.27459)
calibrated <- data.frame(x, y, z)
calibrated <- apply_radians(calibrated)
```

apply_updown	<i>Apply Elevation (UpDown)</i>
--------------	---------------------------------

Description

Apply Elevation (UpDown)

Usage

```
apply_updown(x)
```

Arguments

x Calibrated acceleration data frame.

Value

Measure column appended to end of calibrated data frame.

Examples

```
x <- c(0.14268, 0.21757, -0.529, -0.36383)
y <- c(0.26385, 0.27295, 0.29220, 0.79510)
z <- c(0.27722, 0.20296, 0.35092, 0.27459)
calibrated <- data.frame(x, y, z)
calibrated <- apply_updown(calibrated)
```

binfile_summary	<i>Bin File Summary</i>
-----------------	-------------------------

Description

Bin File Summary

Usage

```
binfile_summary(input, recursive = TRUE)
```

Arguments

input	Bin file path - single bin file or folder of bin files.
recursive	TRUE applies the operation to all nested elements.

Details

Wrapper function that calls create_summary for bin files only.

Value

Data frame of bin file or MPI summary.

calc_autocalparams	<i>Calculate Auto-calibration Parameters</i>
--------------------	--

Description

Function to calculate auto-calibration parameters from known still points from a bin file that create a unitary sphere.

Usage

```
calc_autocalparams(
  binfile,
  binfile_path,
  output_folder,
  sphere_points,
  use_temp = TRUE,
  spherecrit = 0.3,
  maxiter = 500,
  tol = 1e-13
)
```

Arguments

binfile	Text lines read from an open connection to a bin file.
binfile_path	Path to the bin file to be processed.
output_folder	Path to the folder containing GENEAcCore run outputs and Measurement Period Information (MPI) files.
sphere_points	List of points that populate a unitary sphere and their associated temperature in the form (x,y,z,temp).
use_temp	Allows auto-calibration to be run with and without temperature compensation.
spherecrit	The minimum required acceleration value for each axis in both directions for auto-calibration to be reliable.
maxiter	The maximum number of sphere fit iterations attempted during auto-calibration.
tol	The limit of incremental sphere fit improvements before auto-calibration is considered complete.

Value

List of auto-calibration parameters within the measurement period information (MPI).

Examples

```
binfile_path <- system.file("extdata/10Hz_calibration_file_20Nov25.bin", package = "GENEAcCore")
output_folder <- tempdir()
con <- file(binfile_path, "r")
binfile <- readLines(con, skipNul = TRUE)
close(con)
MPI <- create_MPI(binfile, binfile_path, output_folder)
nonmovement_list <- detect_nonmovement(binfile, binfile_path, output_folder)
MPI <- calc_autocalparams(
  binfile, binfile_path, output_folder,
  nonmovement_list$non_movement$sphere_points
)
```

check_MPI

Print out a summary of MPI file(s)

Description

Function to print out a summary of an MPI (Measurement Process Information) file, or all MPI files found if a directory path is supplied.

Usage

```
check_MPI(input, print_output = TRUE)
```

Arguments

input	Fully qualified path to a MPI file or a directory containing MPI files, the directory is searched recursively.
print_output	Boolean to print MPI summary to console.

Examples

```
## Not run:
check_MPI("fully qualified path to a directory structure containing MPI files")
check_MPI("fully qualified path to a MPI file")

## End(Not run)
```

create_event_mapping *Create Event Mapping*

Description

Create Event Mapping

Usage

```
create_event_mapping(events, start_time, end_time, max_row_number)
```

Arguments

events	Data frame containing the start and end index of each event.
start_time	Name of the column in events containing the start index of the events.
end_time	Name of the column in events containing the end index of the events.
max_row_number	Number of rows in the source vector the events describe

Details

Enumerate a vector to identify which event each measurement belongs to.

Value

List of mapped events.

Examples

```
events <- data.frame(
  "start" = c(1, 5, 10, 15),
  "end" = c(4, 9, 14, 19)
)
time_series <- rnorm(25)
period_number <- create_event_mapping(events, "start", "end", length(time_series))
```

create_MPI	<i>Create Measurement Period Information</i>
------------	--

Description

Create Measurement Period Information

Usage

```
create_MPI(binfile, binfile_path, output_folder, out_rds = TRUE)
```

Arguments

binfile	Text lines read from an open connection to a bin file.
binfile_path	Path to the bin file to be processed.
output_folder	Folder to write MPI file in.
out_rds	Allows RDS output to be saved during MPI creation.

Details

Function to create measurement period information (MPI) from a GENEActiv bin file

Value

List of measurement period information.

Examples

```
binfile_path <- system.file("extdata/20Hz_file.bin", package = "GENEActiv")
con <- file(binfile_path, "r")
binfile <- readLines(con, skipNul = TRUE)
close(con)
output_folder <- tempdir()
MPI <- create_MPI(binfile, binfile_path, output_folder, out_rds = FALSE)
```

create_summary	<i>Create Summary</i>
----------------	-----------------------

Description

Create Summary

Usage

```
create_summary(input, path_type, recursive)
```

Arguments

input	Input type of either a bin file path, MPI path or an MPI object.
path_type	The file type within the folder to create summary for.
recursive	TRUE applies the operation to all nested elements.

Details

Function to create a summary of key information of a bin file or MPI path.

Value

Data frame of bin file or MPI summary.

detect_nonmovement	<i>Detect Non-movement</i>
--------------------	----------------------------

Description

Detect Non-movement

Usage

```
detect_nonmovement(  
  binfile,  
  binfile_path,  
  output_folder,  
  still_seconds = 120,  
  sd_threshold = 0.013,  
  temp_seconds = 240,  
  border_seconds = 300,  
  long_still_seconds = 120 * 60,  
  delta_temp_threshold = -0.7,  
  posture_changes_max = 2,  
  non_move_duration_max = 12 * 60 * 60  
)
```

Arguments

binfile	Text lines read from an open connection to a bin file.
binfile_path	Path to the bin file to be processed.
output_folder	Path to the folder containing GENEAcCore run outputs and Measurement Period Information (MPI) files.
still_seconds	The number of seconds included in the rolling standard deviation calculation for stillness to determine the shortest detection duration.
sd_threshold	The threshold applied to the rolling standard deviation of combined acceleration to determine stillness.

temp_seconds The number of seconds included in the rolling temperature difference calculation or non-wear which also determines the shortest detection duration.

border_seconds The minimum number of seconds of a still event to be classed as a new bout.

long_still_seconds The minimum number of seconds of a still bout that is classed as non-wear.

delta_temp_threshold The threshold applied to the rolling temperature difference to determine non-wear.

posture_changes_max The maximum number of adjoining events that make up a single bout.

non_move_duration_max The maximum number of seconds of a still bout to be classed as non-movement. Still bouts with a duration longer than this number is automatically classed as non-wear.

Details

Function to detect non-movement bouts, non-wear events and points in a 1Hz downsampled bin file.

Value

List of sphere points, non-movement bouts and non-wear events.

Examples

```
binfile_path <- system.file("extdata/20Hz_file.bin", package = "GENEAcore")
con <- file(binfile_path, "r")
binfile <- readLines(con, skipNul = TRUE)
close(con)
output_folder <- tempdir()
MPI <- create_MPI(binfile, binfile_path, output_folder)
MPI <- detect_nonmovement(binfile, binfile_path, output_folder)
```

detect_transitions	<i>Detect Transitions</i>
--------------------	---------------------------

Description

Detect Transitions

Usage

```
detect_transitions(
  binfile,
  binfile_path,
  output_folder,
  minimum_event_duration = 5,
```



```

    x_cpt_penalty = 18,
    y_cpt_penalty = 25,
    z_cpt_penalty = 16,
    cut_time_24hr = "15:00"
  )

```

Arguments

<code>binfile</code>	Text lines read from an open connection to a bin file.
<code>binfile_path</code>	Path to the bin file to be processed.
<code>output_folder</code>	Path to the folder containing GENEAcCore run outputs and Measurement Period Information (MPI) files.
<code>minimum_event_duration</code>	The minimum interval between changepoint transitions.
<code>x_cpt_penalty</code>	The manual penalty value applied in the PELT changepoint algorithm for the x axis, see cpt.var .
<code>y_cpt_penalty</code>	The manual penalty value applied in the PELT changepoint algorithm for the y axis, see cpt.var .
<code>z_cpt_penalty</code>	The manual penalty value applied in the PELT changepoint algorithm for the z axis, see cpt.var .
<code>cut_time_24hr</code>	Time in 24h to split the days up by.

Details

Function to detect mean and variance changepoints in 1Hz acceleration data from a bin file.

Value

List of time, index and day number of each transition within the measurement period information.

Examples

```

binfile_path <- system.file("extdata/20Hz_file.bin", package = "GENEAcCore")
con <- file(binfile_path, "r")
binfile <- readLines(con, skipNul = TRUE)
close(con)
output_folder <- tempdir()
MPI <- create_MPI(binfile, binfile_path, output_folder)
MPI <- detect_transitions(binfile, binfile_path, output_folder)

```

geneacore

*Main GENEAcCore Function***Description**

Main GENEAcCore Function which performs the following tasks:

- Checks for and reads or creates Measurement Processing Information (MPI)
- Downsamples the file to 1Hz and detects periods of non movement
- Calculates auto calibration settings for the device
- Identifies transitions (if processing events)
- Applies auto calibration scaling
- Counts steps
- Performs aggregations by event or epoch
- Writes out data

Usage

```
geneacore(
  data_folder = data_folder,
  cut_time_24hr = "15:00",
  output_epochs = TRUE,
  epoch_duration = 1,
  output_events = TRUE,
  output_steps = FALSE,
  output_csv = FALSE,
  timer = FALSE,
  summary = FALSE
)
```

Arguments

<code>data_folder</code>	Folder that contains raw data bin files to process or path to single bin file.
<code>cut_time_24hr</code>	Time in 24h to split the days up by.
<code>output_epochs</code>	Create epoch outputs.
<code>epoch_duration</code>	Specify duration of fixed epochs.
<code>output_events</code>	Create event outputs.
<code>output_steps</code>	Include step counts and stepping rate outputs.
<code>output_csv</code>	Allows CSV output to be saved during epoch and event processing.
<code>timer</code>	Print elapsed times of each process.
<code>summary</code>	Create bin file summary for all files in data folder.

Value

RDS and CSV of Measurement Period Information, Epoch measures and Event measures.

`get_UniqueBinFileIdentifier`*Generate Unique Bin File Identifier*

Description

Generate Unique Bin File Identifier

Usage

```
get_UniqueBinFileIdentifier(binfile)
```

Arguments

`binfile` Text lines read from an open connection to a bin file.

Details

Function to create a UniqueBinFileIdentifier from a GENEActiv bin file.

Value

Single string identifier.

Examples

```
binfile_path <- system.file("extdata/20Hz_file.bin", package = "GENEAcore")
con <- file(binfile_path, "r")
binfile <- readLines(con, skipNul = TRUE)
close(con)
UniqueBinFileIdentifier <- get_UniqueBinFileIdentifier(binfile)
```

`get_UniqueBinFileIdentifier.dir`*Generate Unique Bin File Identifier for each bin file within a directory path*

Description

Generate Unique Bin File Identifier for each bin file within a directory path

Usage

```
get_UniqueBinFileIdentifier.dir(binfile_directory)
```

Arguments

binfile_directory
Path to the file to be processed

Details

Function to create a UniqueBinFileIdentifier from a GENEActiv bin file.

Value

Named vector, the vector name is the filename and the value the identifier.

Examples

```
binfile_directory <- system.file("extdata", package = "GENEAcCore")
UniqueBinFileIdentifiers <- get_UniqueBinFileIdentifier.dir(binfile_directory)
```

```
get_UniqueBinFileIdentifier.filepath
```

Generate Unique Bin File Identifier from a file path

Description

Generate Unique Bin File Identifier from a file path

Usage

```
get_UniqueBinFileIdentifier.filepath(binfile_path)
```

Arguments

binfile_path Path to the file to be processed

Details

Function to create a UniqueBinFileIdentifier from a GENEActiv bin file.

Value

Single string identifier.

Examples

```
binfile_path <- system.file("extdata/20Hz_file.bin", package = "GENEAcCore")
UniqueBinFileIdentifier <- get_UniqueBinFileIdentifier.filepath(binfile_path)
```

get_UniqueBinFileIdentifier.vector

Generate Unique Bin File Identifier

Description

Generate Unique Bin File Identifier

Usage

```
get_UniqueBinFileIdentifier.vector(binfile)
```

Arguments

binfile Text lines read from an open connection to a bin file.

Details

Function to create a UniqueBinFileIdentifier from a GENEActiv bin file.

Value

Single string identifier.

Examples

```
binfile_path <- system.file("extdata/20Hz_file.bin", package = "GENEAcore")
con <- file(binfile_path, "r")
binfile <- readLines(con, skipNul = TRUE)
close(con)
UniqueBinFileIdentifier <- get_UniqueBinFileIdentifier(binfile)
```

MPI_summary

MPI Summary

Description

MPI Summary

Usage

```
MPI_summary(input, recursive = TRUE)
```

Arguments

input MPI path - single MPI file or folder of MPI files.
recursive TRUE applies the operation to all nested elements.

Details

Wrapper function that calls create_summary for MPI only.

Value

Data frame of MPI summary.

```
remove_short_transitions
```

Remove Short Transitions

Description

Remove Short Transitions

Usage

```
remove_short_transitions(cpts_var_df, cut_times, minimum_event_duration)
```

Arguments

cpts_var_df Calculated changepoints data frame.
 cut_times Start of day transitions data frame.
 minimum_event_duration The minimum interval between changepoint transitions.

Details

Identify and remove transitions shorter than the minimum event duration.

Value

Data frame of all transitions including start of day with short transitions removed.

Examples

```
changepoints <- data.frame(
  time = c(1677855218, 1677855598, 1677855661, 1677855679),
  index = c(86019, 86399, 62, 80),
  day = c(1, 1, 2, 2)
)
cut_times <- data.frame(time = 1677855600, index = 1, day = 2)
transitions <- remove_short_transitions(changepoints, cut_times, 5)
```

sample_binfile	<i>Sample Bin File</i>
----------------	------------------------

Description

Sample Bin File

Usage

```
sample_binfile(  
    binfile,  
    binfile_path,  
    output_folder,  
    start_time = NULL,  
    end_time = NULL,  
    downsample = TRUE,  
    output_csv = FALSE,  
    save_raw = FALSE  
)
```

Arguments

binfile	Text lines read from an open connection to a bin file.
binfile_path	Path to the bin file to be processed.
output_folder	Path to the folder containing GENEActiv run outputs and Measurement Period Information (MPI) files.
start_time	Time stamp to start the read from, default start of file.
end_time	Time stamp to end the read from, default end of file.
downsample	Logical to determine whether to downsample the file, default TRUE.
output_csv	Allow outputs of bin file sampling to be saved as CSV.
save_raw	Save daily raw sampled data as RDS to for quicker reprocessing, default FALSE.

Details

Function to read in a GENEActiv bin file with option to downsample to 1Hz. This can read the whole of the file or just a portion of it by setting the start_time and end_time parameters.

Value

List of 1Hz downsampled data or raw sample data.

Examples

```
binfile_path <- system.file("extdata/20Hz_file.bin", package = "GENEAcore")
output_folder <- tempdir()
con <- file(binfile_path, "r")
binfile <- readLines(con, skipNul = TRUE)
close(con)
measurements <- sample_binfile(binfile, binfile_path, output_folder)
```

step_counter

Step Counter

Description

Function to calculate the number and variance of the steps in the data.

Usage

```
step_counter(
  step_data,
  sample_frequency = 100,
  filter_order = 2,
  boundaries = c(0.5, 5),
  Rp = 3,
  hysteresis = 0.05,
  fun = c("GENEAcount", "mean", "sd", "stepdiff")
)

stepCounter(...)
```

Arguments

step_data	The data to use for calculating the steps. This should be a vector of acceleration values.
sample_frequency	The sampling frequency of the data, in hertz, when calculating the step number (default 100).
filter_order	single integer, order of the Chebyshev bandpass filter, passed to argument n of cheby1 .
boundaries	length 2 numeric vector specifying lower and upper bounds of Chebychev filter (default c(0.5, 5) Hz), passed to argument W of butter or cheby1 .
Rp	the decibel level that the cheby filter takes, see cheby1 .
hysteresis	The hysteresis applied after zero crossing. (default 100mg)
fun	character vector naming functions by which to summarize steps. "count" is an internally implemented summarizing function that returns step count.
...	Additional arguments passed to internal aggregation functions.

Value

Returns a vector with length fun.

Examples

```
d1 <- sin(seq(0.1, 100, 0.1)) / 2 + rnorm(1000) / 10 + 1
Steps4 <- step_counter(d1)
length(Steps4)
mean(Steps4)
sd(Steps4)
plot(Steps4)
```

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